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## Enzim Restriksi Endonuklease

# Kemampuan Akhir yang Diharapkan

- Mahasiswa dapat menjelaskan prinsip kerja enzim restriksi endonuklease
- Mahasiswa dapat menjelaskan pengelompokan enzim restriksi
- Mahasiswa dapat menjelaskan macam-macam enzim restriksi pada vektor



# Restriction Enzymes Mapping

# What are restriction enzymes?

- Molecular scissors that cut double stranded DNA molecules at specific points.
- Found naturally in a wide variety of prokaryotes
- An important tool for manipulating DNA.

# Discovery

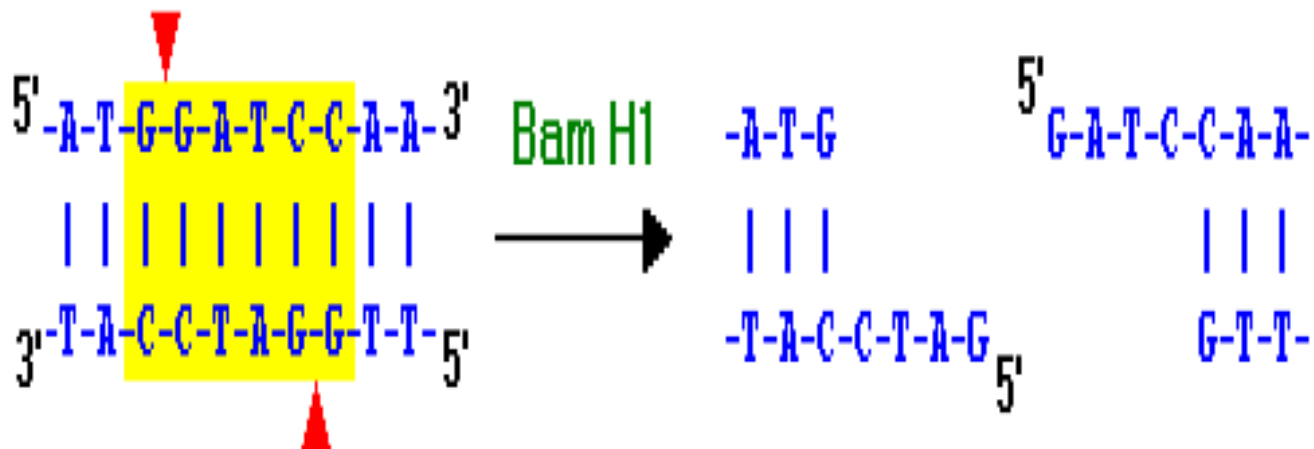
- Arbor and Dussoix in 1962 discovered that certain bacteria contain Endonucleases which have the ability to cleave DNA.
- In 1970 Smith and colleagues purified and characterized the cleavage site of a Restriction Enzyme.
- Werner Arbor, Hamilton Smith and Daniel Nathans shared the 1978 Nobel prize for Medicine and Physiology for their discovery of Restriction Enzymes.

# ENZYMES USED IN MOLECULAR BIOLOGY

Restriction enzymes	Cut the DNA
<u>Alkaline phosphatase</u>	Removes phosphate groups from 5' ends of DNA (prevents unwanted re-ligation of cut DNA)
<u>DNA ligase</u>	Joins compatible ends of DNA fragments (blunt/blunt or complementary cohesive ends). Uses ATP
<u>DNA polymerase I</u>	Synthesises DNA complementary to a DNA template in the 5'-to-3'direction. Starts from an oligonucleotide primer with a 3' OH end
<u>Exonuclease III</u>	Digests nucleotides progressively from a DNA strand in the 3' -to-5' direction
<u>Polynucleotide kinase</u>	Adds a phosphate group to the 5' end of double- or single-stranded DNA or RNA. Uses ATP
<u>RNase A</u>	Nuclease which digests RNA, not DNA
<u>Taq DNA polymerase</u>	Heat-stable DNA polymerase isolated from a thermostable microbe ( <i>Thermus aquaticus</i> )

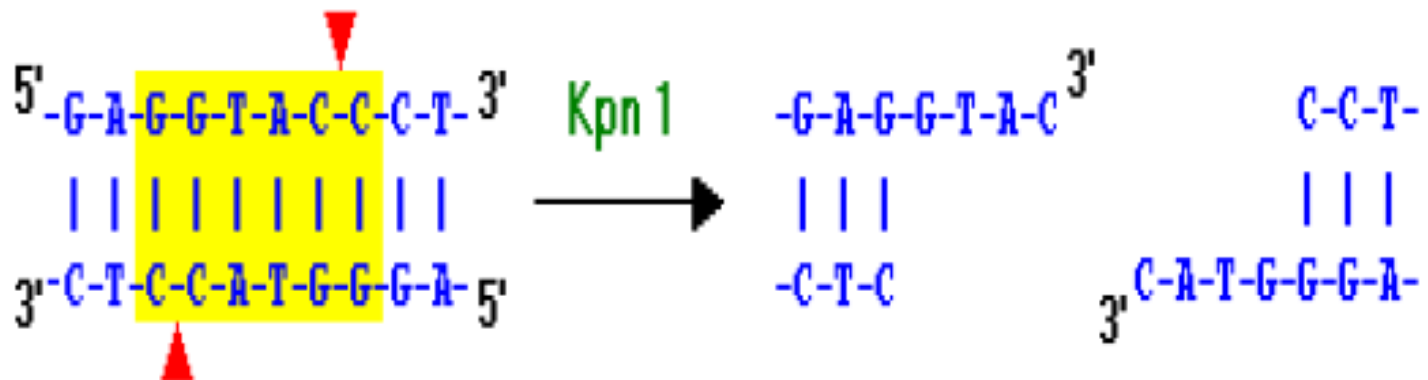
# 5' OVERHANGS

- **5' overhangs:** The enzyme cuts asymmetrically within the recognition site such that a short single-stranded segment extends from the 5' ends. *Bam* HI cuts in this manner.



# 3' OVERHANGS

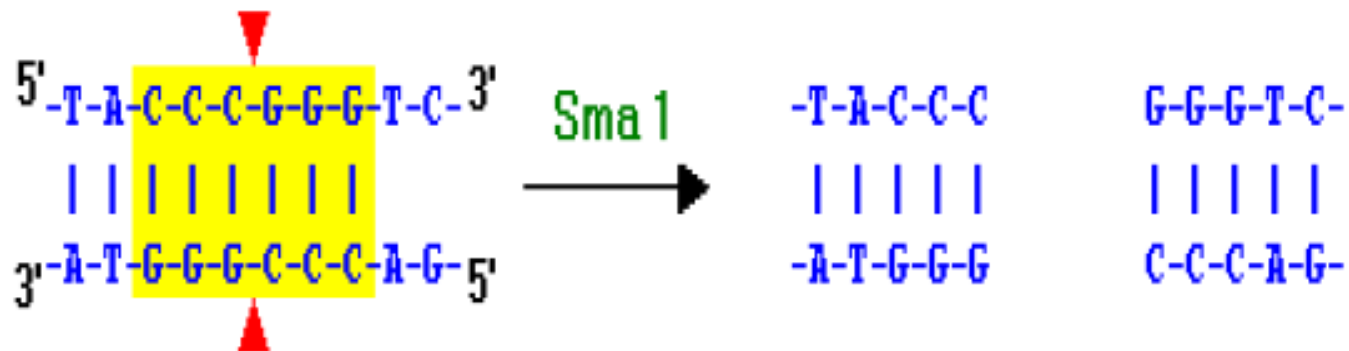
- **3' overhangs:** Again, we see asymmetrical cutting within the recognition site, but the result is a single-stranded overhang from the two 3' ends. *Kpn*I cuts in this manner.





# BLUNT ENDS

- **Blunts:** Enzymes that cut at precisely opposite sites in the two strands of DNA generate blunt ends without overhangs. *Sma* I is an example of an enzyme that generates blunt ends.

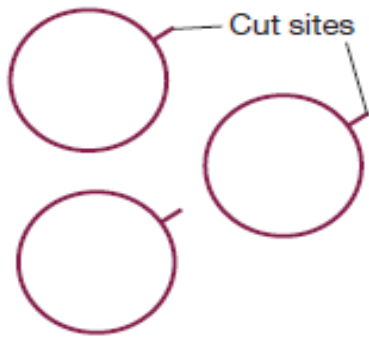


# Types of Restriction Enzymes

	<b>Cleavage site</b>	<b>Location of methylase</b>	<b>Examples</b>
Type I	Random Around 1000bp away from recognition site	Endonuclease and methylase located on a single protein molecule	EcoK I EcoA I CfrA I
Type II	Specific Within the recognition site	Endonuclease and methylase are separate entities	EcoR I BamH I Hind III
Type III	Random 24-26 bp away from recognition site	Endonuclease and methylase located on a single protein molecule	EcoP I Hinf III EcoP15 I

# Pemotongan DNA pada Teknologi DNA Rekombinan

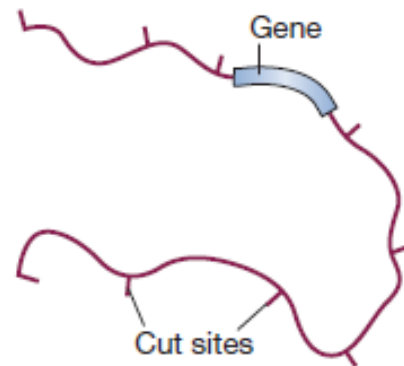
(a) Vector molecules



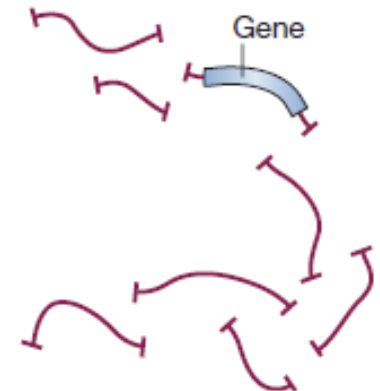
DAN

Each vector molecule must be cut once, each at the same position

(b) The DNA molecule containing the gene to be cloned

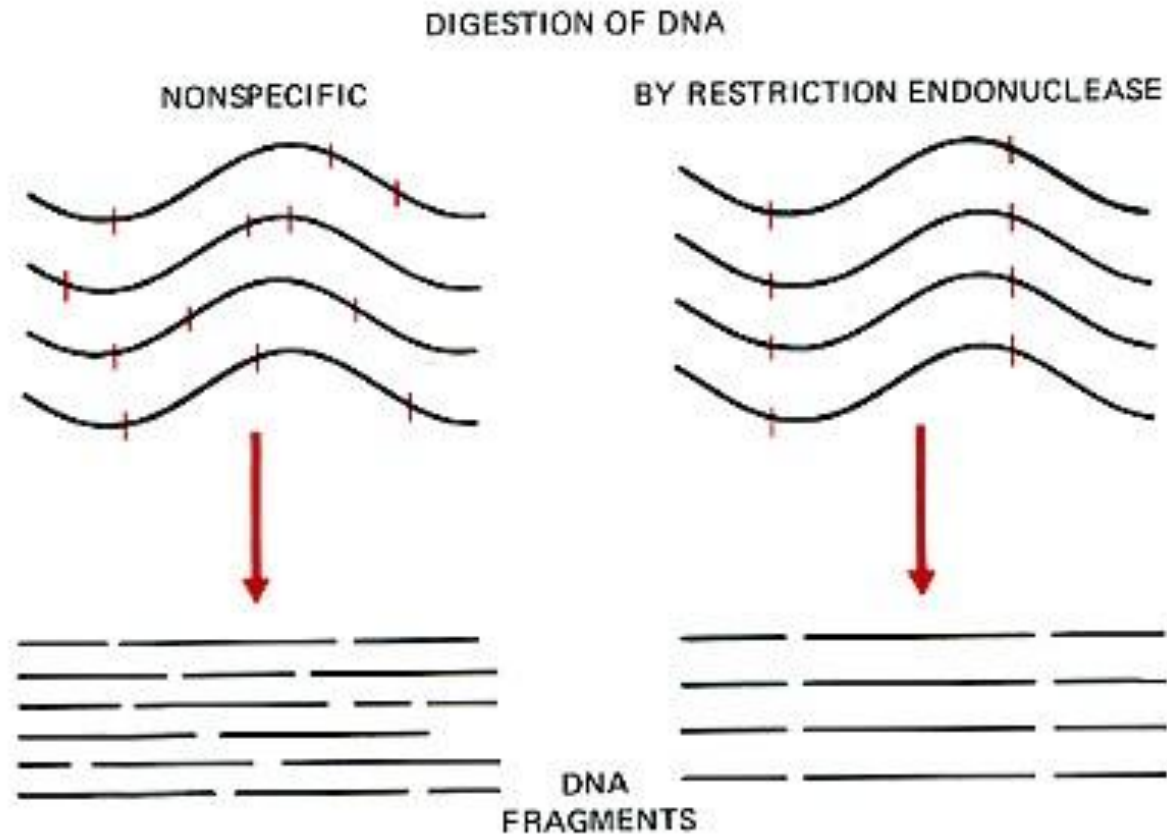


Large DNA molecule



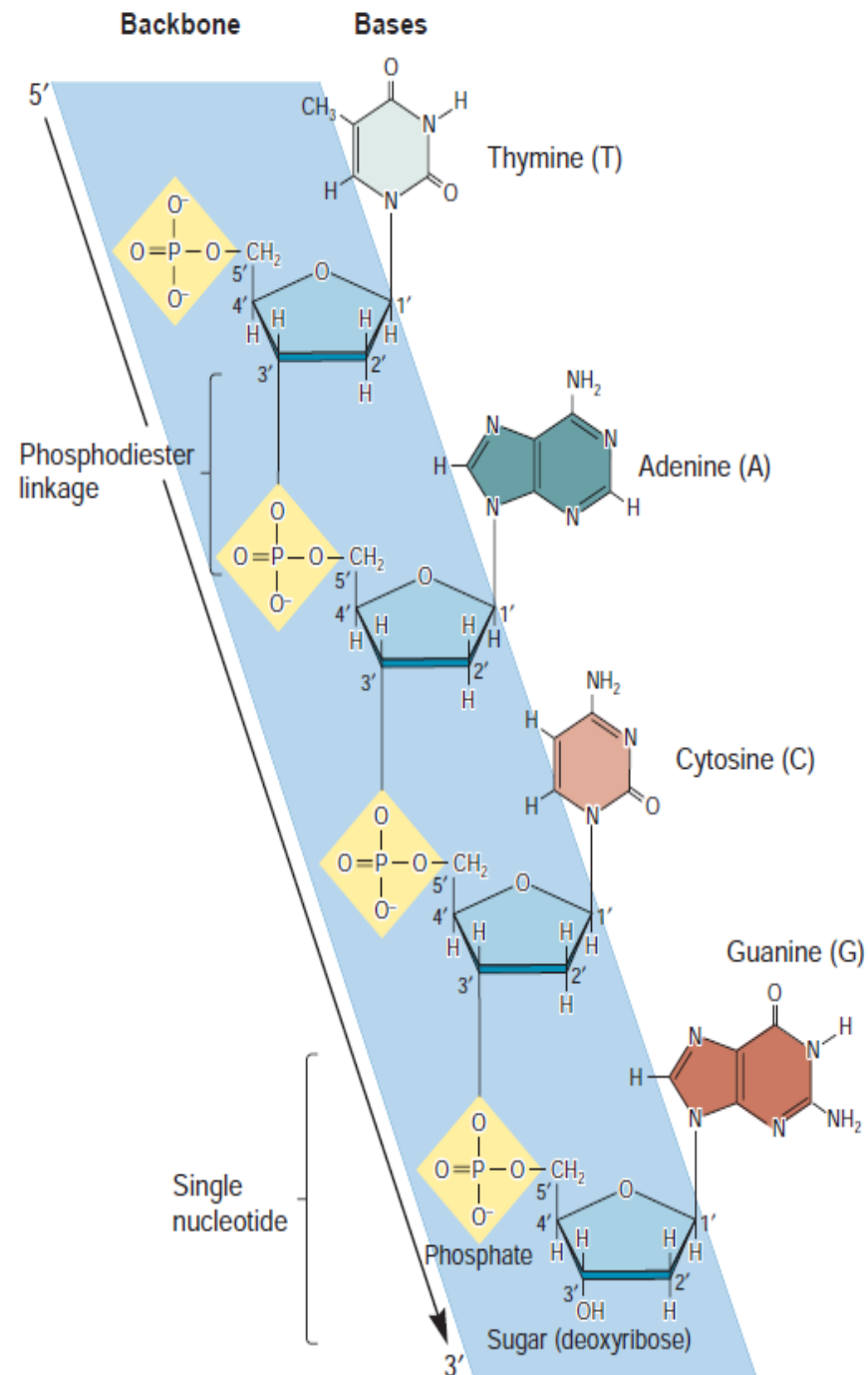
Fragments small enough to be cloned

# Pemotongan DNA pada Teknologi DNA Rekombinan Tidak Acak



# Enzim Restriksi Endonuklease

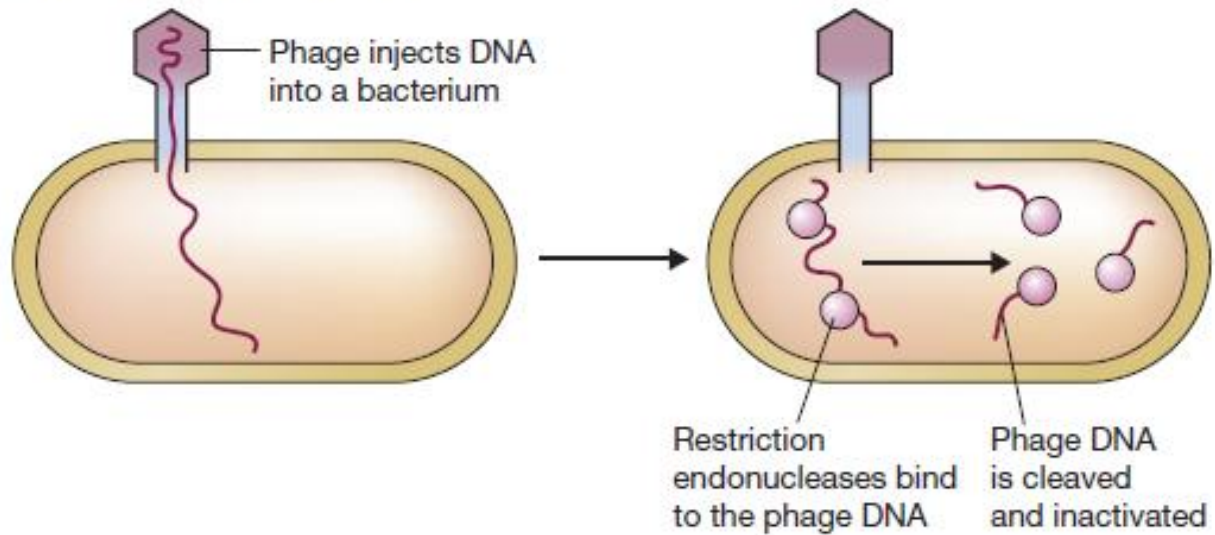
- Dapat memotong DNA → memotong pada ikatan fosfodiester
- Memotong di bagian dalam → **endonuklease**
- Disebut juga dengan **enzim restriksi**
- **Dihasilkan oleh bakteri dan Archaea**



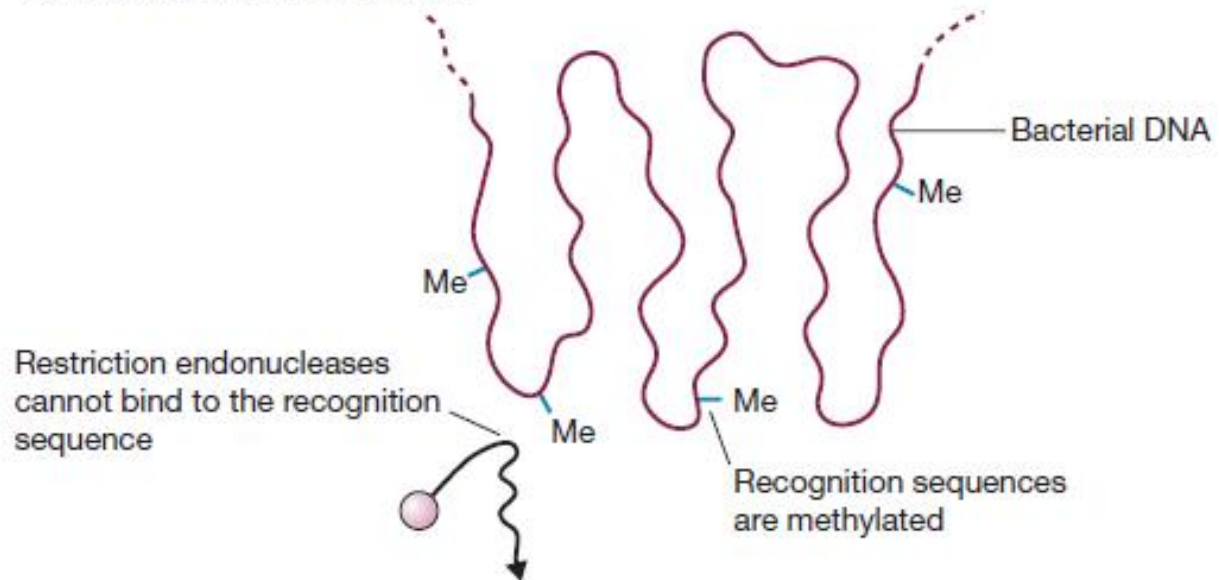
# Sejarah Penemuan Enzim Restriksi

- Beberapa peneliti menemukan bahwa bakteriofaga tidak dapat menginfeksi bakteri
- Hal ini dipelajari lebih lanjut oleh W. Arber, H. Smith dan D. Nathans
- Bakteri menghasilkan enzim yang dapat memotong DNA virus

(a) Restriction of phage DNA



(b) Bacterial DNA is not cleaved



# Sejarah Penemuan Enzim Restriksi

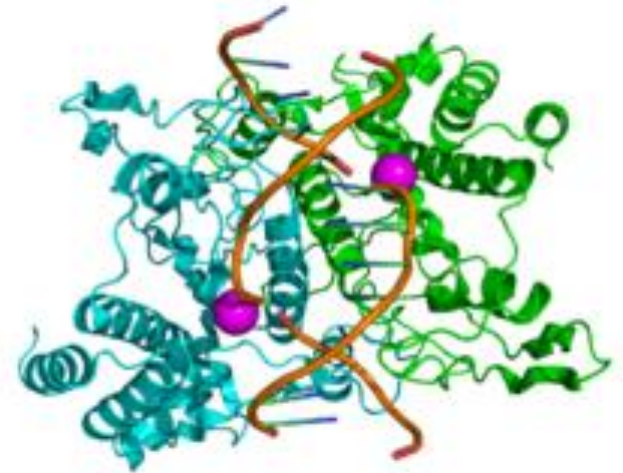
- Menghasilkan enzim restriksi yang pertama yaitu **HindIII**
- Pada tahun 1978 ketiga peneliti tersebut mendapatkan hadiah Nobel dalam Fisiologi atau Kedokteran





# Tipe-tipe Enzim Restriksi Endonuklease

- **Tipe I** → akan memotong DNA di luar daerah pemotongan spesifik, tidak digunakan dalam rekombinasi DNA
- **Tipe II** → memotong DNA pada situs yang spesifik, digunakan dalam rekombinasi DNA
- **Tipe III** → akan memotong DNA di luar daerah pemotongan spesifik, memerlukan 2 sekuen yang sama, tidak digunakan untuk rekombinasi DNA
- **Tipe IV** → akan mengenali DNA dengan metilasi, tidak digunakan dalam rekombinasi DNA



## TYPES AND ACTIVITIES OF RESTRICTION ENZYMES

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### Type I

Cleaves DNA at random sites far from its recognition sequence

### Type II

Cleaves DNA at defined positions close to or within its recognition sequence

### Type IIG

Cleaves outside its recognition sequence with both REase and MTase enzymatic activities in the same protein

### Type IIP

Cleaves symmetric targets and cleavage sites

### Type IIS

Recognizes asymmetric sequences

### Type III

Cleaves outside its recognition sequence and require two sequences in opposite orientations within the same DNA

### Type IV

Cleaves modified (e.g., methylated) DNA

# Penamaan Enzim Restriksi

- Huruf yang digunakan tidak dimiringkan
- Huruf pertama merupakan nama genus bakteri yang menghasilkan
- Huruf kedua dan ketiga merupakan nama spesies bakteri yang menghasilkan
- Huruf keempat merupakan strain dari bakteri yang menghasilkan
- Nomor romawi menunjukkan urutan penemuan enzim tersebut

# Penamaan Enzim Restriksi

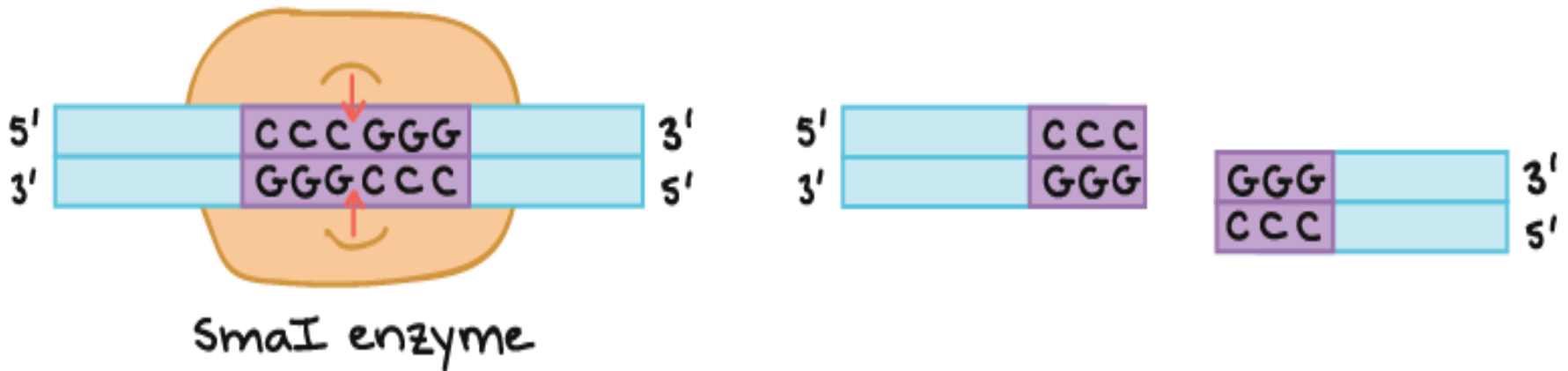
- Contoh : enzim EcoRI
  - Berasal dari *Escherichia coli*
  - strain RY
- Contoh : enzim HindIII
  - Berasal dari *Haemophilus influenza*
  - Serotipe d

# Enzim Restriksi Tipe II Memotong Pada Situs Pemotongan yang Spesifik

ENZYME	ORGANISM	RECOGNITION SEQUENCE*	BLUNT OR STICKY END
<i>EcoRI</i>	<i>Escherichia coli</i>	GAATTC	Sticky
<i>BamHI</i>	<i>Bacillus amyloliquefaciens</i>	GGATCC	Sticky
<i>BglII</i>	<i>Bacillus globigii</i>	AGATCT	Sticky
<i>PvuI</i>	<i>Proteus vulgaris</i>	CGATCG	Sticky
<i>PvuII</i>	<i>Proteus vulgaris</i>	CAGCTG	Blunt
<i>HindIII</i>	<i>Haemophilus influenzae</i> R <sub>d</sub>	AAGCTT	Sticky
<i>HinfI</i>	<i>Haemophilus influenzae</i> R <sub>f</sub>	GANTC	Sticky
<i>Sau3A</i>	<i>Staphylococcus aureus</i>	GATC	Sticky
<i>AluI</i>	<i>Arthrobacter luteus</i>	AGCT	Blunt
<i>TaqI</i>	<i>Thermus aquaticus</i>	TCGA	Sticky
<i>HaeIII</i>	<i>Haemophilus aegyptius</i>	GGCC	Blunt
<i>NotI</i>	<i>Nocardia otitidis-caviarum</i>	GCGGCCGC	Sticky
<i>SfiI</i>	<i>Streptomyces fimbriatus</i>	GGCCNNNNGGCC	Sticky

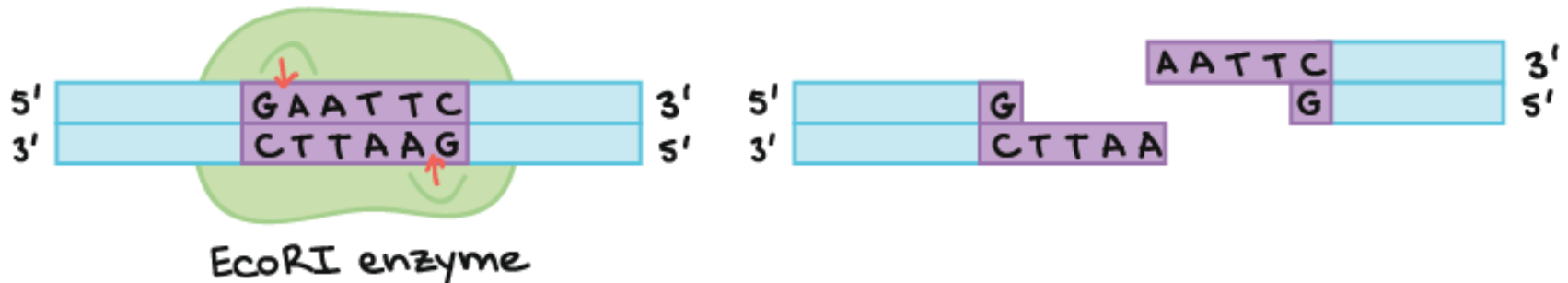
# Hasil Pemotongan Enzim Restriksi

- Ujung-ujung tumpul (*Blunt ends*)



# Hasil Pemotongan Enzim Restriksi

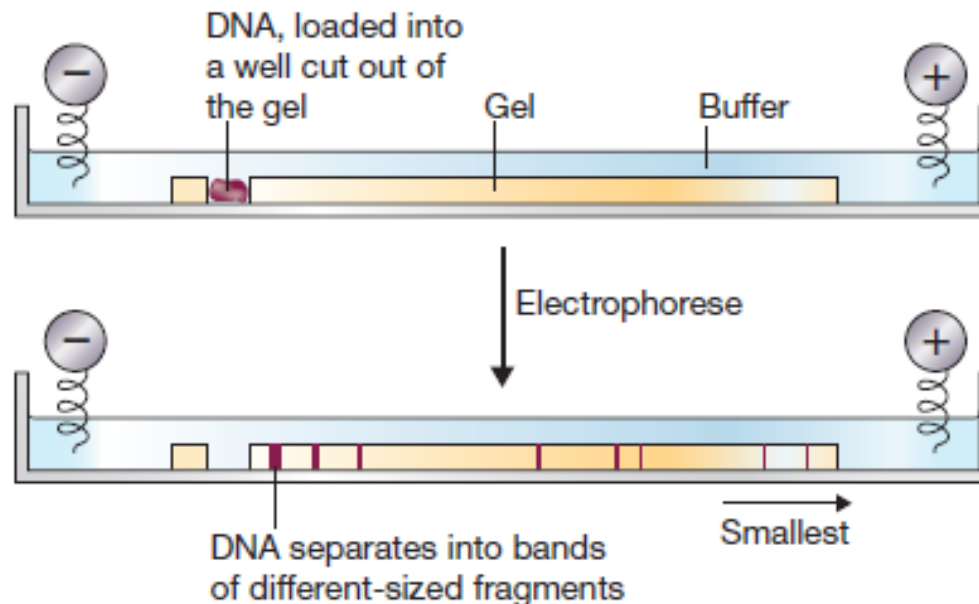
- Ujung-ujung menggantung (*Sticky ends*)



# Analisa Hasil Restriksi DNA

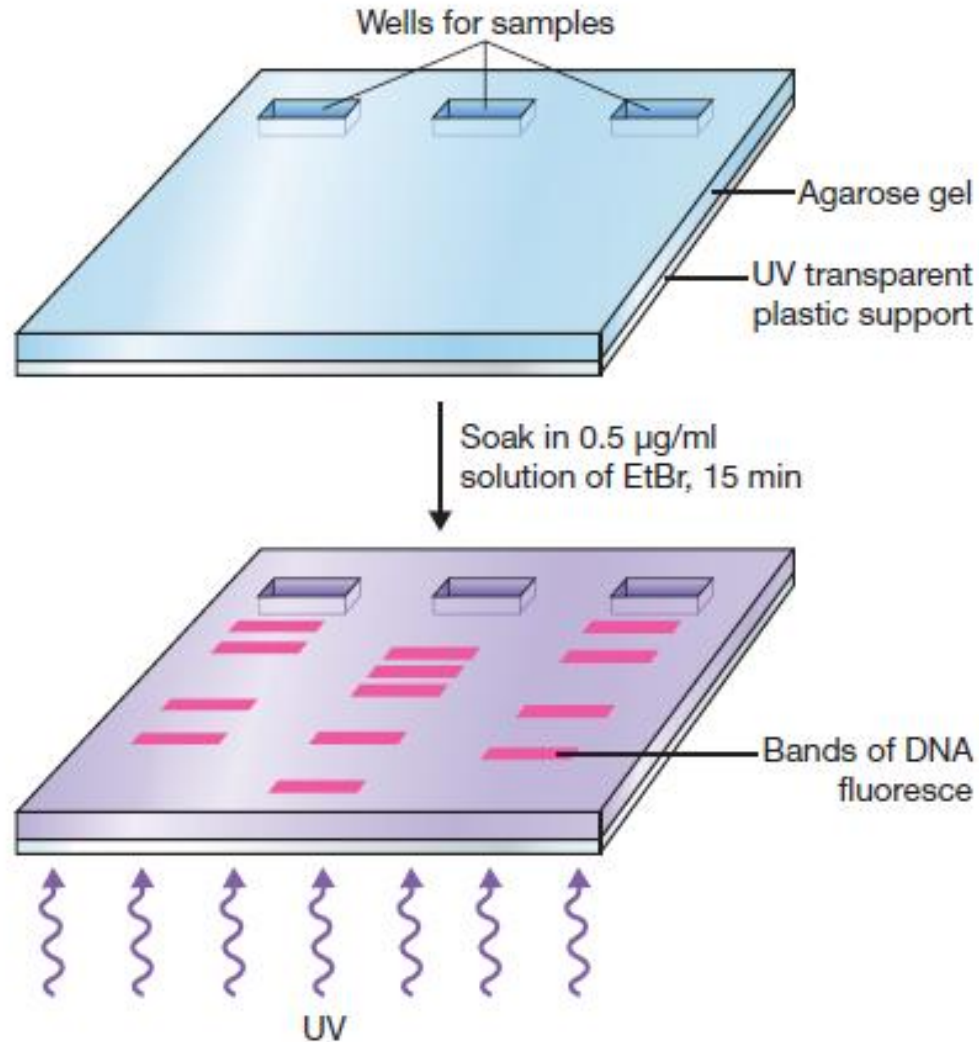
- Dengan elektroforesis gel
- Baik gel agarose maupun poliakrilamid

(b) Gel electrophoresis

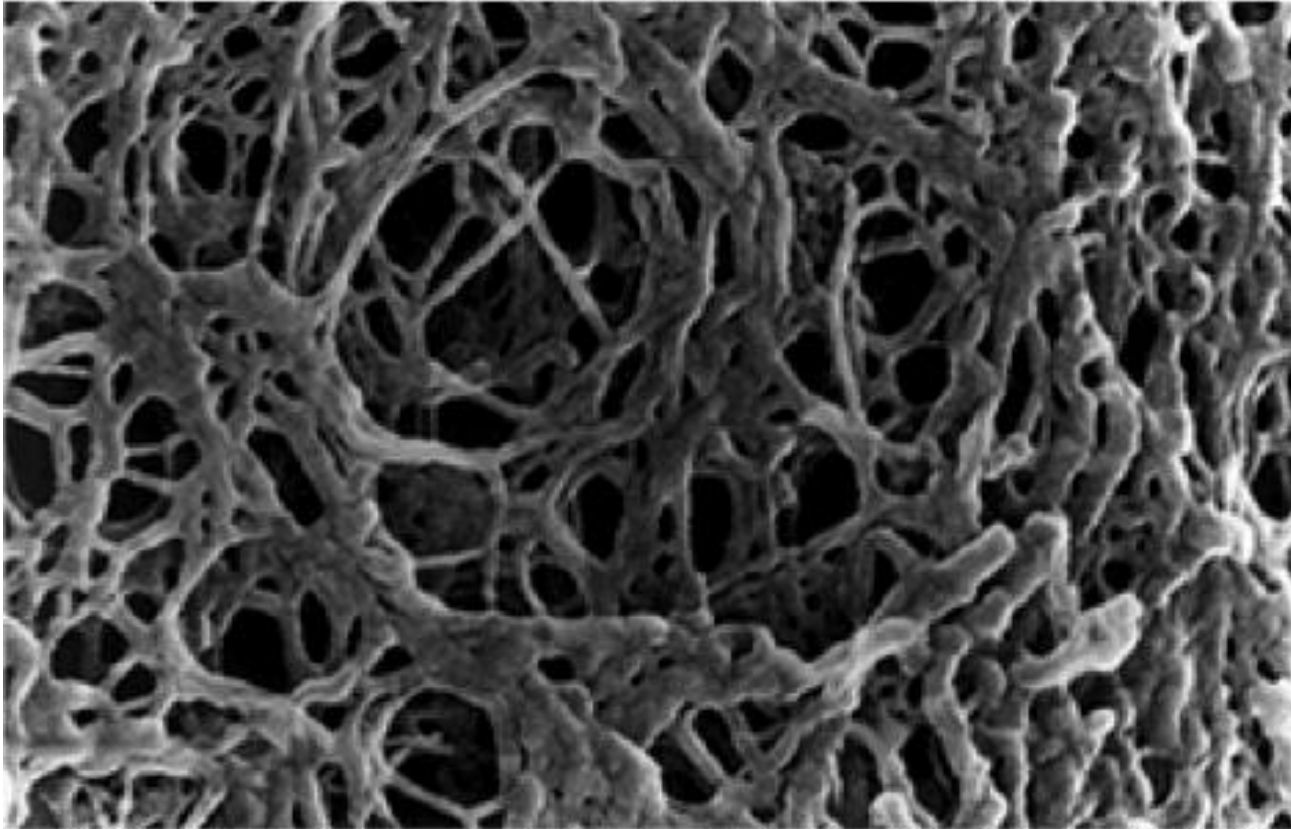




# Elektroforesis Gel Agarose



# Pori-pori pada Gel Agarose dan Poliakrilamid

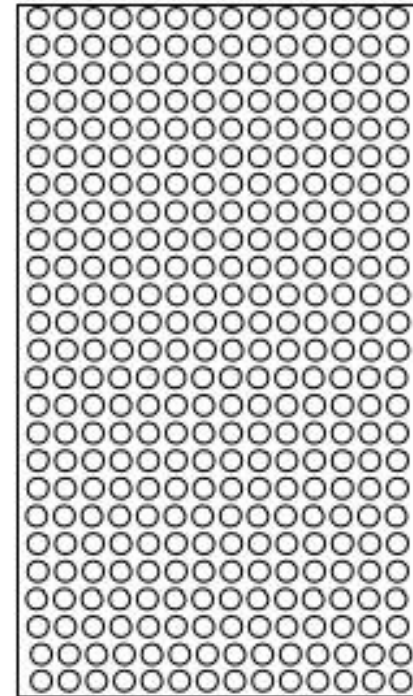
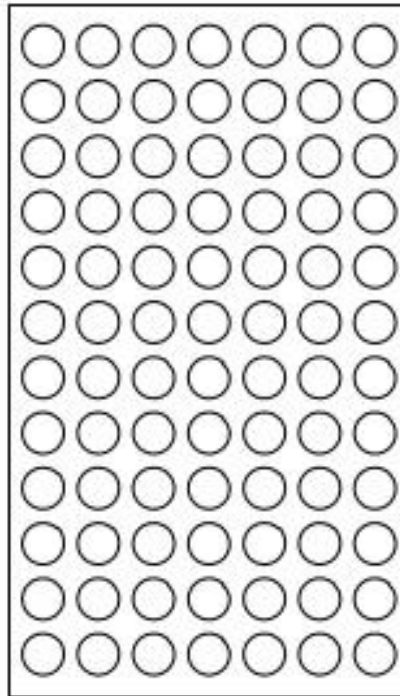
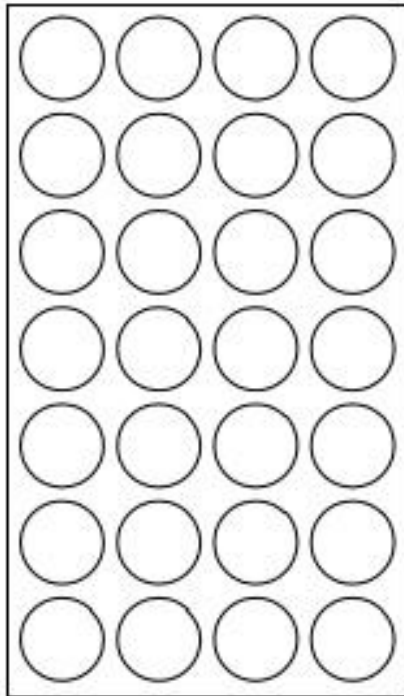


# Agarose concentration

**As we increase the concentration of agarose (g/ml) the pores gets smaller!**



Concentration →



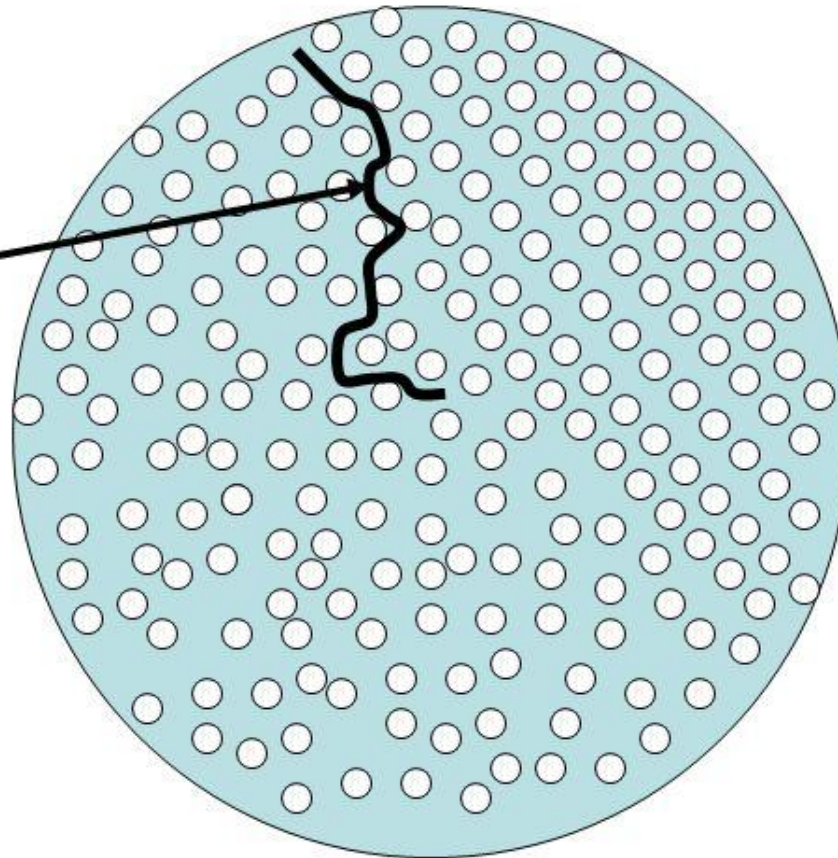
*m*

(<https://www.slideshare.net/hhalhaddad/281-lec30-moltech2>)

# Microscopic view of a gel

- pore
- agarose

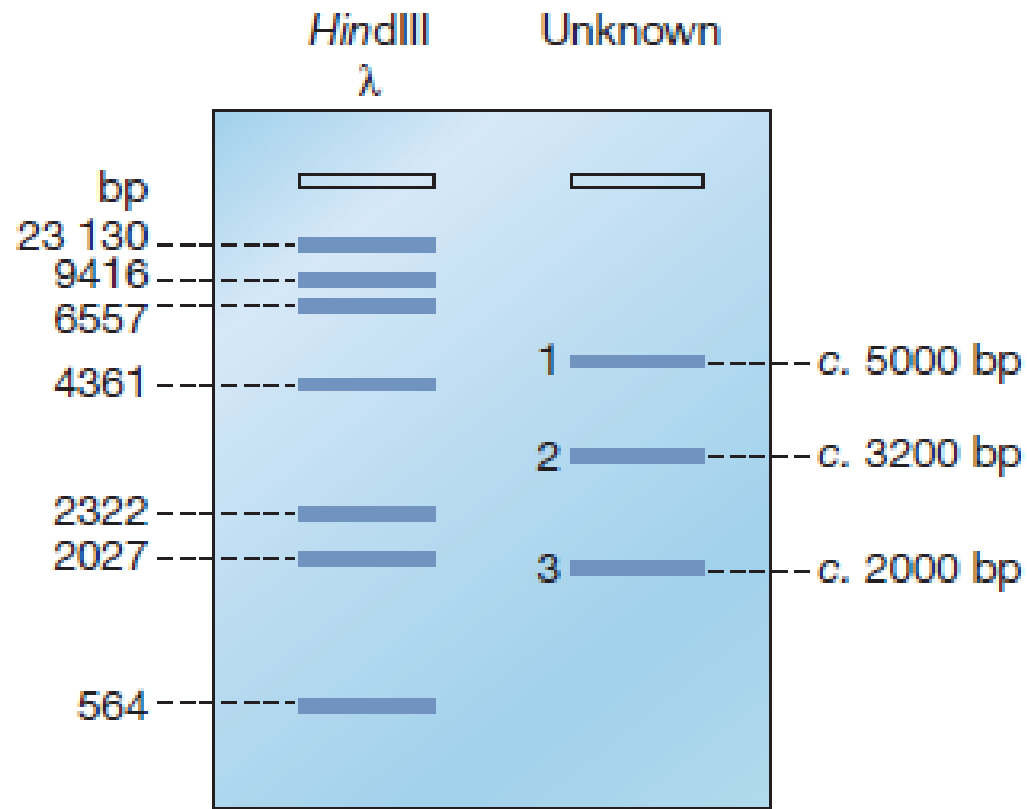
Fragment  
of DNA “running”  
through gel



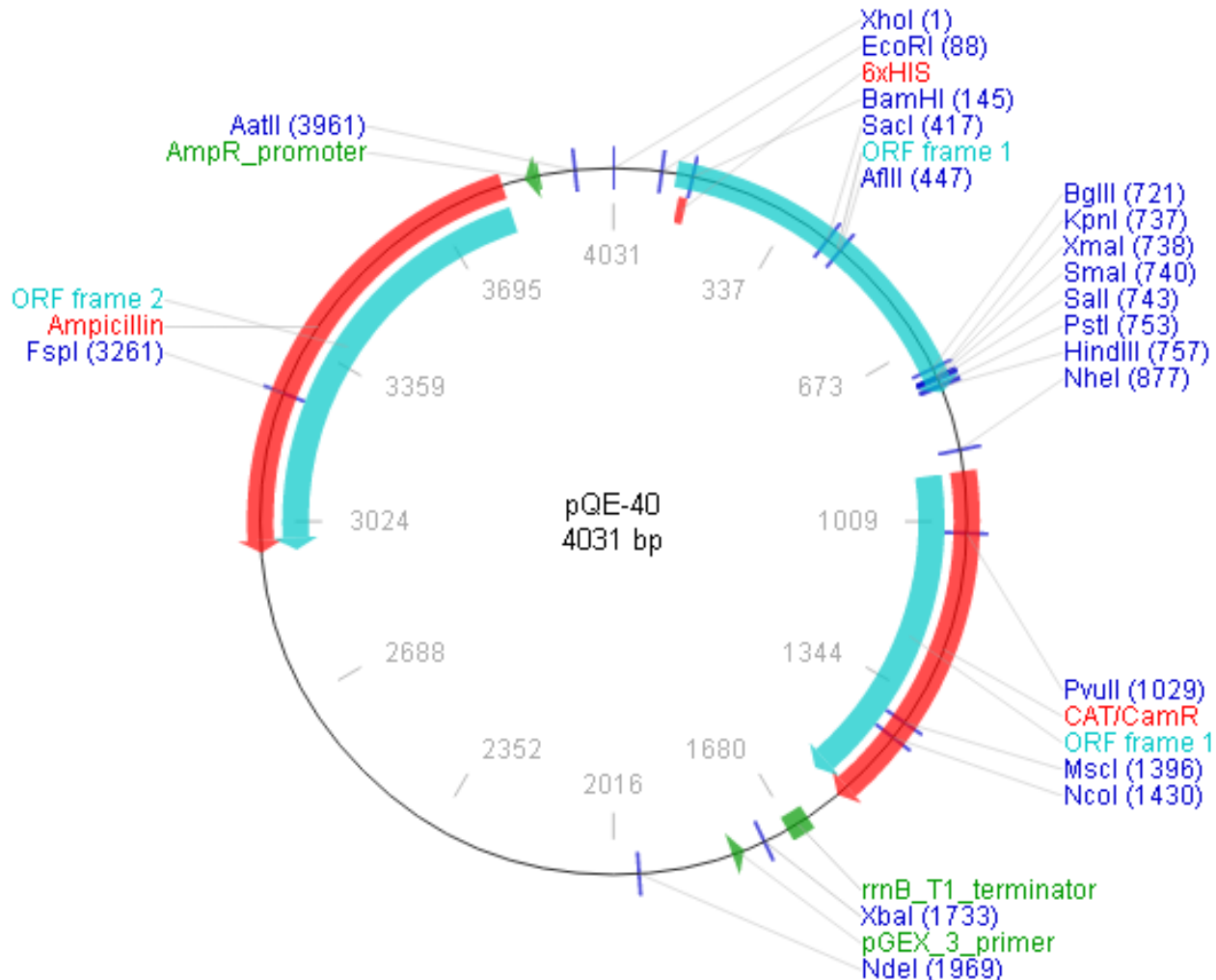
*(<http://slideplayer.com/slide/7876390/>)*

# Analisa Hasil Restriksi

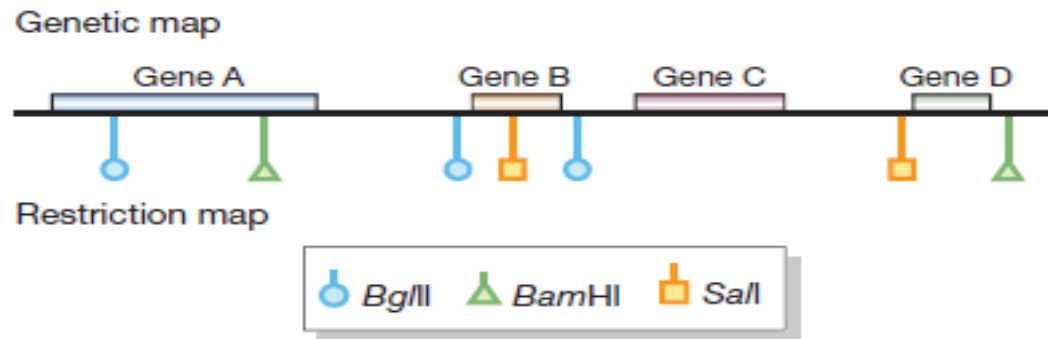
(a) Rough estimation by eye



# Peta Enzim Restriksi pada Vektor



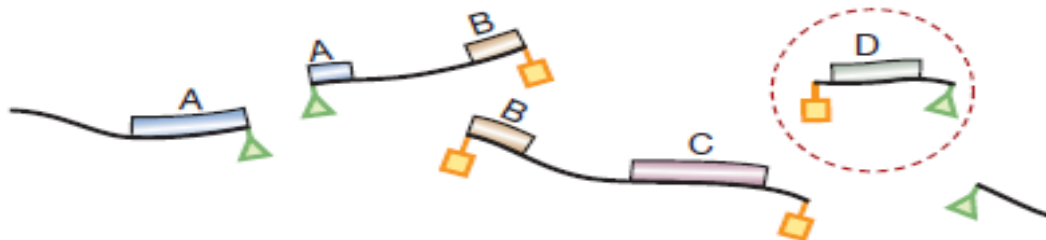
# Cara Mengetahui Situs Restriksi Pada Gen atau Genom



To obtain gene B, digest with *Bgl*II



To obtain gene D, digest with *Bam*HI + *Sal*I



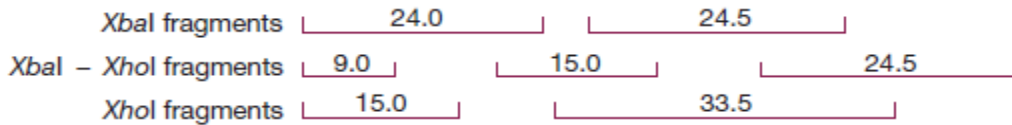
## Single and double digestions

Enzyme	Number of fragments	Sizes (kb)
<i>Xba</i> I	2	24.0, 24.5
<i>Xho</i> I	2	15.0, 33.5
<i>Kpn</i> I	3	1.5, 17.0, 30.0
<i>Xba</i> I + <i>Xho</i> I	3	9.0, 15.0, 24.5
<i>Xba</i> I + <i>Kpn</i> I	4	1.5, 6.0, 17.0, 24.0

### Conclusions:

(1) As  $\lambda$  DNA is linear, the number of restriction sites for each enzyme is *Xba*I 1, *Xho*I 1, *Kpn*I 2.

(2) The *Xba*I and *Xho*I sites can be mapped:



The only possibility is: 15.0 *Xho*I 9.0 *Xba*I 24.5

(3) All the *Kpn*I sites fall in the 24.5 kb *Xba*I fragment, as the 24.0 kb fragment is intact after *Xba*I–*Kpn*I double digestion. The order of the *Kpn*I fragments can be determined only by partial digestion.

### Partial digestion

Enzyme	Fragment sizes (kb)
<i>Kpn</i> I – limiting conditions	1.5, 17.0, 18.5, 30.0, 31.5, 48.5

### Conclusions:

- 48.5 kb fragment = uncut  $\lambda$ .
- 1.5, 17.0 and 30.0 kb fragments are products of complete digestion.
- 18.5 and 31.5 kb fragments are products of partial digestion.

The *Kpn*I map must be: 30.0 *Kpn*I 1.5 17.0

Therefore the complete map is: 15.0 *Xho*I 9.0 *Xba*I 6.0 *Kpn*I 1.5 17.0



**ZAMAN OLD**



# Welcome to RestrictionMapper - on line restriction mapping the easy way.

Maps sites for restriction enzymes, a.k.a. restriction endonucleases, in DNA sequences. Also does virtual digestion.

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Conformation	Include	Sequence Info	Menu
<p>Circular <input type="radio"/></p> <p>Linear <input checked="" type="radio"/></p>	<p>Select Individual Enzymes</p> <p>All Enzymes ▲</p> <p>AarI</p> <p>AasI</p> <p>AatI</p> <p>AatII</p> <p>AccI</p> <p>AcclI ▼</p>	<p>No non-base letters. Numbers and spaces OK.</p> <p>Paste Sequence Here</p> <div style="border: 1px solid black; height: 150px; width: 100%;"></div>	
<p><b>Sort By</b></p> <p>1. frequency ▼</p> <p>2. overhang ▼</p> <p>3. name ▼</p>			<p>Map Sites</p> <p>Virtual Digest</p> <p>Reset Form</p>
<p><b>Filter By</b></p> <p>Maximum Cuts all ▼</p> <p>Minimum Site Length 5 ▼</p>	<p>All Commercial <input checked="" type="radio"/></p> <p>NEB only <input type="radio"/></p> <p>5' overhang <input checked="" type="checkbox"/></p> <p>3' overhang <input checked="" type="checkbox"/></p> <p>blunt <input checked="" type="checkbox"/></p> <p>Prototypes Only <input type="radio"/></p> <p>All Isoschizomers <input type="radio"/></p>	<p>Name your sequence</p> <p>Untitled</p>	

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## GenScript Restriction Enzyme Map Analysis Tools

\*\* This online tool helps you analyze restriction enzyme cutting maps.


Paste in DNA Sequence:

pGAD-C1-SEC13.dna (Circular / 7549 bp)

File Edit View Enzymes Features Primers Actions Window Help

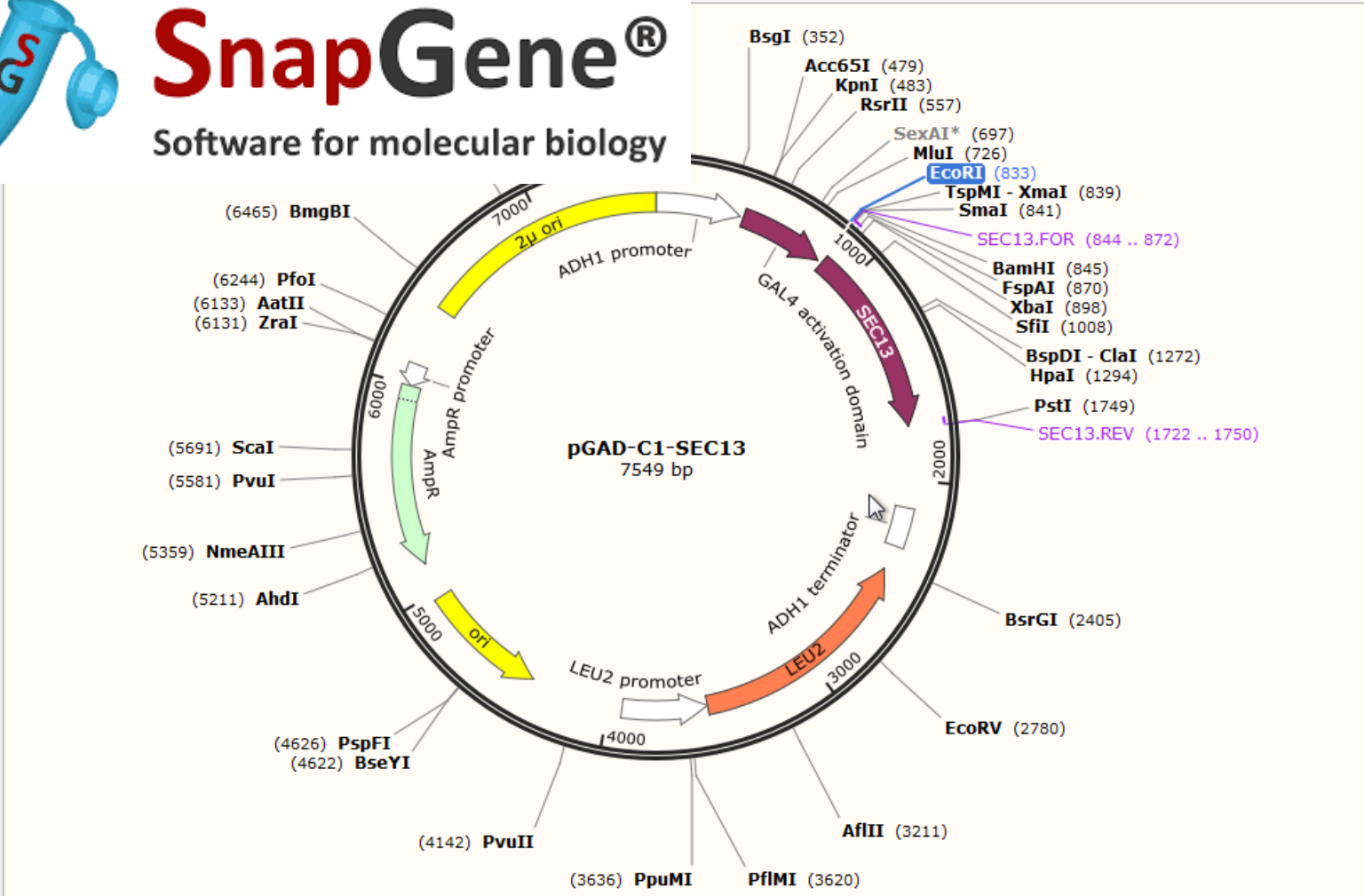
New Open Save Print Undo Redo Copy Paste

7549 bp



# SnapGene®

Software for molecular biology



**pGAD-C1-SEC13**  
7549 bp

Restriction Enzyme Sites (Clockwise from top):

- BsgI (352)
- Acc65I (479)
- KpnI (483)
- RsrII (557)
- SexAI\* (697)
- MluI (726)
- EcoRI (833)
- TspMI - XmaI (839)
- SmaI (841)
- SEC13.FOR (844 .. 872)
- BamHI (845)
- FspAI (870)
- XbaI (898)
- SfiI (1008)
- BspDI - ClaI (1272)
- HpaI (1294)
- PstI (1749)
- SEC13.REV (1722 .. 1750)
- BsrGI (2405)
- EcoRV (2780)
- AflIII (3211)
- PfIMI (3620)
- PpuMI (3636)
- PvuII (4142)
- PspFI (4626)
- BseYI (4622)
- AhdI (5211)
- NmeAIII (5359)
- PvuI (5581)
- ScaI (5691)
- ZraI (6131)
- AatII (6133)
- PfoI (6244)
- BmgBI (6465)

Other Features:

- ADH1 promoter
- ADH1 terminator
- LEU2 promoter
- AmpR promoter
- 2μ ori
- ori
- GAL4 activation domain

Unique 6+ Cutters (Nonredundant)

Map Sequence Enzymes Features Primers History

Description Panel

pGAD-C1-SEC13.dna (Circular / 7549 bp)

File Edit View Enzymes Features Primers Actions Window Help

New Open Save Print Undo Redo Copy Paste

Selected: EcoRI (833) 7549 bp

10 20 30 40 50 60 70 80

MluI

GTATAACGCGTTTGGAACTACTACAGGGATGTTTAATACCACTACAATGGATGATGATATACTATCTATTTCGATGATG  
CATATTGCGCAAACCTTAGTGATGTCCTACAAATTATGGTGATGTTACCTACTACATATATTGATAGATAAGCTACTAC

870 875 880 885 890

Y N A F G I T T G M F N T T M D D V Y N Y L F D D

GAL4 activation domain

SmaI BamHI FspAI

EcoRI XmaI TspMI

SEC13.FOR

GT GGGATCCATGGTCGTCATAGCTAATGCCG

AAGATACCCACCAAACCCAAAAAAGAGATCGAATTCCTCCCGGGGGATCCATGGTCGTCATAGCTAATGCCGACAACGAA  
TTCTATGGGGTGGTTTGGGTTTTTTTCTCTAGCTTAAGGGGGCCCCCTAGGTACCAGCAGTATCGATTACGCGTGTGCTT

895 900

E D T P P N P K K E I E F P G G S M V V I A N A H N E

GAL4 activation domain SEC13

XbaI

TTAATCCATGACGCTGTTCTAGACTATTATGGGAAGCGCCTTGCAACCTGCTCTTCTGACAAGACAATCAAGATCTTTGA  
AATTAGGTACTGCGACAAGATCTGATAATACCCTTCGCGGAACGTTGGACGAGAAGACTGTTCTGTTAGTTCTAGAACT

15 20 25 30 35

L I H D A V L D Y Y G K R L A T C S S D K T I K I F E

SEC13

SfiI

AGTCGAAGGAGAAACACACAAGTTAATAGACACGTTGACTGGCCACGAAGGCCAGTTTGGCGTGTGATTGGGCACATC

1040

Unique 6+ Cutters (Nonredundant)

Map Sequence Enzymes Features Primers History

Description Panel

pGAD-C1-SEC13.dna (Circular / 7549 bp)

File Edit View Enzymes Features Primers Actions Window Help

New Open Save Print Undo Redo Copy Paste

Selected: **AvaI (839)** 7549 bp

Chosen Enzymes: 6+ Cutters (198) from "Nonredundant Commercial"

Enzyme	Sites	Numbers	Lines
(69)	0	<i>69 chosen enzymes do not cut</i>	
<b>AatII</b>	1	6133	
AccI	3	227 2053 2189	
<b>Acc65I</b>	1	479	
AclI	3	621 5437 5810	
AcuI	5	2555 4866 5878 6568 6635	
AfeI	9	594 6513 6591 6610 7249 7311 7374 7436 7499	
<b>AflII</b>	1	3211	
AflIII	4	726 990 2789 4318	
<b>AhdI</b>	1	5211	
AjuI	6	1416 1448 3406 3438 7123 7155	
AlwNI	2	1744 4734	
ApaLI	4	1807 4632 5878 6375	
ApoI	16	252 583 833 1043 1604 1909 1956 2066 2133 2194 2227 2327 2384 2889 3804 3819	
AseI	7	311 437 881 1930 4089 4148 5383	
<b>AvaI</b>	3	443 <b>839</b> 7253	
BaeI	2	3905 3938	
BaeGI	5	1036 1811 4636 5882 6379	
<b>BamHI</b>	1	845	
BanI	3	479 4062 5159	
BanII	2	1174 1312	
BarI	4	218 250 6566 6598	
BbsI	3	95 3074 6623	
BcgI	4	1262 1296 5716 5750	
BciVI	2	4521 6048	
BdaI	6	1718 1752 4064 4098 4389 4423	

\* Methylation-sensitive sites

AvaI 3 443 **839** 7253  
 ... 396 6414 739

Map Sequence Enzymes Features Primers History Description Panel

pGAD-C1-SEC13.dna (Circular / 7549 bp)

File Edit View Enzymes Features Primers Actions Window Help

New Open Save Print Undo Redo Copy Paste

Selected: **AvaI (839)** 7549 bp

Chosen Enzymes: 6+ Cutters (198) from "Nonredundant Commercial"

Enzyme	Sites	Numbers	Lines
(69)	0	69 chosen enzymes do not cut	
<b>AatII</b>	1		
AccI	3		
<b>Acc65I</b>	1		
AdI	3		
AcuI	5		
AfeI	9		
<b>AflII</b>	1		
AflIII	4		
<b>AhdI</b>	1		
AjuI	6		
AlwNI	2		
ApaLI	4		
ApoI	16		
AseI	7		
<b>AvaI</b>	3		
BaeI	2		
BaeGI	5		
<b>BamHI</b>	1		
BanI	3		
BanII	2		
BarI	4		
BbsI	3		
BcgI	4		
BciVI	2		
BdaI	6		

\* Methylation-sensitive sites Noncutters

AvaI 3

Map Sequence Enzymes Features Primers History Description Panel